

**SAKI SULTANA**

**B. Pharm (Pharmacy), University of Dhaka, 2006**

**M. Pharm (Pharmacy), University of Dhaka, 2007**

**DEPARTMENT OF BIOCHEMISTRY AND MOLECULAR  
BIOLOGY**

**TITLE OF  
THESIS:** GLYCOPHINGOLIPID METABOLISM AND  
THE GLYCOPHINGOLIPID-  
METABOLIZING ENZYME BETA-  
GLUCOSIDASE-2: BIOCHEMICAL AND  
CELL BIOLOGICAL STUDIES

**TIME/DATE:** 1:30 pm, Monday, December 2, 2019

**PLACE:** Room 3107, The Mona Campbell Building, 1459  
LeMarchant Street

**EXAMINING COMMITTEE:**

Dr. Timothy E. Shutt, Department of Medical Genetics, University of  
Calgary (External Examiner)

Dr. Barbara Karten, Department of Biochemistry and Molecular Biology,  
Dalhousie University (Reader)

Dr. Stefan Krueger, Department of Physiology and Biophysics, Dalhousie  
University (Reader)

Dr. Petra Kienesberger, Department of Biochemistry and Molecular  
Biology, Dalhousie University (Reader)

Dr. Aarnoud C. van der Spoel, Department of Pediatrics/Biochemistry and  
Molecular Biology, Dalhousie University (Supervisor)

**DEPARTMENTAL  
REPRESENTATIVE:** Dr. Jan Rainey, Department of Biochemistry  
and Molecular Biology, Dalhousie University

**CHAIR:** Dr. Richard Nowakowski, PhD Defence Panel,  
Faculty of Graduate Studies

**ABSTRACT**

Glycosphingolipids (GSLs) are constituents of eukaryotic cell membranes and consist of a lipid (ceramide) linked to one or more sugar residues. GSLs occur in many structurally distinct forms, varying in both their lipid and oligosaccharide domains. Most cells contain multiple structurally different GSLs. Different cell types and animal tissues have distinct and consistent GSL complements, indicating that cellular GSL homeostasis is strictly regulated. Deviations in GSL homeostasis have pathological consequences. Our understanding of the regulation of GSL levels is very limited. In the first part of my project I attempted to assess the kinetics of GSL turnover by testing a mathematical model correlating the rates of GSL biosynthesis and degradation, using genetic and pharmacological approaches to target GSL-metabolizing enzymes. This project, however, produced inconsistent results.

In the second part of my project I studied the involvement of a GSL-metabolizing enzyme in a neurodegenerative disorder. The gene encoding the enzyme  $\beta$ -glucosidase 2 (GBA2) is mutated in patients affected with a form of hereditary spastic paraplegia, SPG46 (SPastic Gait locus #46). These patients present with a combination of spastic paraplegia and cerebellar ataxia and suffer from muscle weakness, and spasticity in the upper and lower limbs along with other neurological symptoms. Currently, the cascade of events leading from mutations in the *GBA2* gene to SPG46 is largely unexplored. I characterized five nonsense and five missense GBA2 mutants and found that all ten mutants are catalytically inactive. The lack of enzyme activity was not associated with decreased protein expression. Native gel electrophoresis showed that, relative to wild-type GBA2, all mutants fold into aberrant conformations, with the exception of the clinically mild Gly683Arg mutant. In addition, C-terminally truncated GBA2 mutants migrate to mitochondria, cause mitochondrial depolarization, and interfere with mitochondrial fusion, resulting in fragmentation of the mitochondrial network. I also identified an internal domain responsible for the mitochondrial import of the truncated GBA2 mutants. Altogether my research provides new insights into the biochemical and cell biological bases of the neurodegenerative disorder SPG46.